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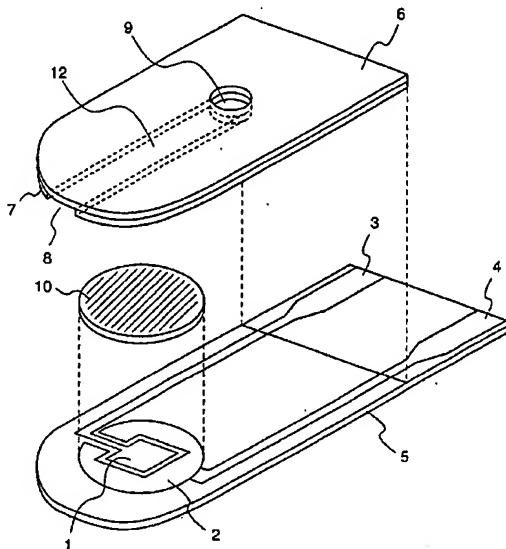
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(54) BIOSENSOR

(57) An electrode layer comprising a working electrode 1 and a counter electrode 2, and a reagent layer 10 are formed on an insulating support 5 and, further, a spacer 7 having a long and narrow cut-out portion on the reagent layer 10 is bonded to a cover 6 having an air hole 9 to form a cavity 12 that sucks blood as a liquid sample by capillary phenomenon, and a portion of side walls of the spacer 7 and the cover 6, which side walls face the cavity 12, is subjected to a treatment for making the portion itself have hydrophilicity.

In the biosensor constructed as described above, when blood is sucked from the cavity 12 by capillary phenomenon, the suction is promoted, and the performance of the sensor is improved. Further, the process of manufacturing the sensor is simplified, resulting in increased productivity.

Fig.1



Description**TECHNICAL FIELD**

5 [0001] The present invention relates to a biosensor for analyzing a specific component in a liquid sample and, more particularly, to a biosensor having a cavity into which a liquid sample is drawn by capillary phenomenon.

BACKGROUND ART

10 [0002] As a biosensor for analyzing a specific component in a liquid sample, there is, for example, a biosensor for detecting a blood sugar level or the like by measuring a current value obtained by a reaction between glucose in blood and a reagent such as glucose oxidase or the like which is held in the sensor.

[0003] Figure 4 is an exploded perspective view illustrating a conventional biosensor for measuring a blood sugar level as described above.

15 [0004] In figure 4, a working electrode 1 and a counter electrode 2 are formed by printing on an insulating support 5 comprising polyethylene terephthalate or the like, and a reagent layer 10 including glucose oxidase and an electron acceptor is formed on these electrodes and, further, a surfactant layer 11 comprising yolk lecithin or the like is formed on the reagent layer 10.

20 [0005] Furthermore, on the surfactant layer 11, a spacer 7 having a long and narrow cut-out portion on the electrodes and the reagent layer 10, and a cover 6 having an air hole are bonded together onto the insulating support 5, to form a cavity 12 in which a specific amount of blood sampled is made to react with the reagent layer 10, and a current value generated by the reaction is detected with the electrodes.

25 [0006] In the biosensor constructed as described above, blood is drawn from a suction inlet 8 into the cavity 12 by capillary phenomenon, and guided to the position where the electrodes and the reagent layer 10 are present. Then, a current value generated by a reaction between the blood and the reagent on the electrodes is read by an external measuring apparatus (not shown) that is connected to the biosensor through leads 3 and 4, and a blood sugar level in the blood is obtained according to the current value.

30 [0007] Conventionally, when blood is applied onto the suction inlet 8 and sampled, in order to draw the blood quickly and deep into the cavity 12 by capillary phenomenon, there has been devised that the surfactant layer 11 is spread so as to cover the reagent layer 10.

35 [0008] However, in the conventional biosensor which facilitates drawing of blood into the cavity 12 by providing the surfactant layer 11 over the reagent layer 10, since the blood is drawn into the cavity while dissolving the surfactant layer 11 and, further, the blood reacts with the reagent layer 10 on the electrodes while dissolving the reagent layer 10, the surfactant layer 11 prevents the reagent layer 10 from dissolving into the blood, and this causes variations in the sensitivity of the sensor or in the measured value, resulting in a detrimental effect on the performance of the sensor.

40 [0009] Further, in the construction of the conventional biosensor, after the reagent layer 10 is formed by spreading a solution including a reagent and an electron acceptor over the electrodes and then drying the solution, formation of the surfactant layer 11 on the reagent layer 10 requires a step of applying and spreading a solution including a surfactant so as to cover the reagent layer 10, and a step of drying the surfactant layer. Therefore, the process of manufacturing the biosensor takes much time, resulting in poor productivity.

45 [0010] The present invention is made to solve the above-described problems and has for its object to provide a biosensor that can promote the flow of blood into the cavity to quickly and sufficiently draw the blood into the cavity, without forming a surfactant layer on the reagent layer.

DISCLOSURE OF THE INVENTION

50 [0011] According to Claim 1 of the present invention, in a biosensor which is provided with a cavity into which a liquid sample is drawn by capillary phenomenon and is able to analyze a component in the liquid sample by a reaction between the drawn liquid sample and a reagent, the surface itself of at least a portion of side walls of the sensor, said side walls facing the cavity, has hydrophilicity.

55 [0012] According to the biosensor constructed as described above, since at least a portion of the side walls of the sensor, which side walls face the cavity into which the liquid sample is drawn by capillary phenomenon, has hydrophilicity at its surface, suction of the liquid sample can be promoted without providing a surfactant layer on the reagent that reacts with the liquid sample. Accordingly, the process of manufacturing the sensor can be simplified.

[0013] According to Claim 2 of the present invention, in the biosensor defined in Claim 1, the side walls of the sensor facing the cavity are made of a resin material in which a surfactant is mixed.

[0014] According to the biosensor constructed as described above, since the side walls having hydrophilicity are made of a resin material in which a surfactant is mixed, suction of the liquid sample can be promoted without providing

a surfactant layer on the reagent that reacts with the liquid sample, and the process of manufacturing the sensor can be simplified.

[0015] According to Claim 3 of the present invention, in the biosensor defined in Claim 2, the amount of the surfactant to be mixed is 0.01 weight % or more.

5 [0016] According to the biosensor constructed as described above, since the side walls of the sensor facing the cavity are made of a resin material into which a surfactant of 0.01 weight % or more is mixed, sufficient blood suction promoting effect can be achieved.

[0017] According to Claim 4 of the present invention, in the biosensor defined in Claim 1, the side walls of the sensor facing the cavity are made of a film the surface of which is covered with a surfactant.

10 [0018] According to the biosensor constructed as described above, since the side walls of the sensor having hydrophilicity are made of a film the surface of which is covered with a surfactant, suction of the liquid sample can be promoted without providing a surfactant layer on the reagent that reacts with the liquid sample and, accordingly, the process of manufacturing the sensor can be simplified.

[0019] According to Claim 5 of the present invention, in the biosensor defined in Claim 1, the side walls of the sensor facing the cavity are made of a film the surface of which is covered with a resin having a hydrophilic polar group.

[0020] According to the biosensor constructed as described above, since the side walls of the sensor having hydrophilicity are made of a film the surface of which is covered with a resin having a hydrophilic polar group, suction of the liquid sample can be promoted without providing a surfactant layer on the reagent that reacts with the liquid sample and, accordingly, the process of manufacturing the sensor can be simplified.

20 [0021] According to Claim 6 of the present invention, in the biosensor defined in Claim 4 or 5, the thickness of the surfactant or the resin having a hydrophilic polar group, which covers the film, is several tens of angstroms or more.

[0022] According to the biosensor constructed as described above, since the side walls of the sensor facing the cavity are made of a film that is covered with the surfactant or the resin having a hydrophilic polar group, sufficient blood suction promoting effect can be achieved.

25 [0023] According to Claim 7 of the present invention, in the biosensor defined in Claim 1, the surface of at least a portion of the side walls forming the cavity is chemically reformed.

[0024] According to the biosensor constructed as described above, since the surface of at least a portion of the side walls forming the cavity is chemically reformed to form the side walls of the sensor having hydrophilicity, suction of the liquid sample can be promoted without providing a surfactant layer on the reagent that reacts with the liquid sample, and accordingly, the process of manufacturing the sensor can be simplified.

30 [0025] According to Claim 8 of the present invention, in the biosensor defined in Claim 7, a hydrophilic functional group is formed on the surface of at least a portion of the side walls facing the cavity, by subjecting the surface to any of the following treatments: plasma discharge, coupling reaction, ozone treatment, and UV treatment.

[0026] According to the biosensor constructed as described above, the surface of at least a portion of the side walls forming the cavity is subjected to any of the following chemical surface treatments: plasma discharge, coupling reaction, ozone treatment, and UV treatment, thereby forming a hydrophilic functional group on the surface. Therefore, the surface of at least a portion of the side walls facing the cavity can have hydrophilicity.

35 [0027] According to Claim 9 of the present invention, in the biosensor defined in Claim 1, the surface of at least a portion of the side walls facing the cavity is made of a rough surface.

40 [0028] According to the biosensor constructed as described above, since the surface of at least a portion of the side walls forming the cavity is roughened to form the side walls of the sensor having hydrophilicity, suction of the liquid sample can be promoted without providing a surfactant layer on the reagent that reacts with the liquid sample, and accordingly, the process of manufacturing the sensor can be simplified.

45 [0029] According to Claim 10 of the present invention, in the biosensor defined in Claim 9, a rough surface is formed at the surface of at least a portion of the side walls facing the cavity, by subjecting the surface to any of the following treatments: sand blasting, electric discharge, non-glare treatment, mat treatment, and chemical plating.

[0030] According to the biosensor constructed as described above, the surface of at least a portion of the side walls forming the cavity is subjected to any of the following treatments: sand blasting, electric discharge, non-glare treatment, mat treatment, and chemical plating, thereby forming a rough surface. Therefore, the surface of at least a portion of the side walls facing the cavity can have hydrophilicity.

50 [0031] According to Claim 11 of the present invention, in the biosensor defined in any of Claims 1 to 10, the surface of the support, on which the reagent that reacts with the liquid sample is formed, also has hydrophilicity.

[0032] According to the biosensor constructed as described above, not only the surface of at least a portion of the side walls forming the cavity but also the surface of the support on which the reagent that reacts with the liquid sample is formed, have hydrophilicity. Therefore, the area of the portion having hydrophilicity in the side walls facing the cavity is increased, whereby the liquid sample can be drawn with higher efficiency.

55 [0033] According to Claim 12 of the present invention, in the biosensor defined in any of Claims 1 to 10, the surface of the support, on which electrodes that detect the reaction between the liquid sample and the reagent are formed,

also has hydrophilicity.

[0034] According to the biosensor constructed as described above, not only the surface of at least a portion of the side walls forming the cavity but also the surface of the support on which the electrodes for detecting the reaction between the liquid sample and the reagent are formed, have hydrophilicity. Therefore, the adhesion of the electrodes to the support on which the electrodes are formed is improved, and the problem of electrode peeling is solved, whereby the reliability of the sensor is improved.

[0035] According to Claim 13 of the present invention, in the biosensor defined in Claim 12, the surface of the support is made of a rough surface, and the level of the rough surface to be formed is 0.001 μm to 1 μm .

[0036] According to the biosensor constructed as described above, since a rough surface having unevenness in a level from 0.001 μm to 1 μm is formed at the surface of at least a portion of the side walls of the sensor facing the cavity, the adhesion is improved.

BRIEF DESCRIPTION OF THE DRAWINGS

[0037] Figure 1 is an exploded perspective view illustrating a biosensor for measuring a blood sugar level, according to embodiments of the present invention.

[0038] Figure 2 is a graph showing the result of a comparison of the sensitivities to blood between a sensor according to Example 1 of the invention and a conventional sensor.

[0039] Figure 3 is a graph showing the result of a comparison of the sensitivities to blood between a sensor according to Example 2 of the present invention and a conventional sensor.

[0040] Figure 4 is an exploded perspective view illustrating a conventional biosensor for measuring a blood sugar level.

BEST MODE TO EXECUTE THE INVENTION

Embodiment 1.

[0041] Hereinafter, a first embodiment of the present invention will be described with reference to figure 1.

[0042] Initially, the construction of a biosensor according to the first embodiment will be described with reference to figure 1.

[0043] Figure 1 is an exploded perspective view of a biosensor according to the first embodiment of the present invention, and this biosensor is different from the conventional one in that the surfactant layer 11 formed on the reaction reagent layer 10 is dispensed with and, as a substitute for the surfactant layer 11, at least a portion of the side walls facing the cavity 12 into which blood is drawn, i.e., at least a portion of parts of the spacer 7 and the cover 6, which parts face the cavity 12, is made to have hydrophilicity by itself, to promote drawing of the blood.

[0044] Hereinafter, a description will be given of specific methods for making the surfaces of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity.

[0045] One of the methods is as follows. An insulating film is formed by mixing a chemical having surface activity such as a surfactant or the like into a material such as polyethylene terephthalate, polycarbonate or the like, and the cover 6 and the spacer 7 are constituted by the insulating film. Thereby, the wettability of the side walls of the cavity 12 is increased, and the blood sampled from the suction inlet 8 can be quickly and reliably drawn into the cavity 12.

[0046] The kinds of surfactants which can be expected to have the above-mentioned effects when being mixed into the insulating film (classified as hydrophilic groups) are as follows: anionic surfactants such as carboxylate, sulfonate, carboxylate, ester phosphate, and the like; cationic surfactants such as primary amine salt, secondary amine salt, tertiary amine salt, quaternary ammonium salt, and the like; ampholytic surfactants such as amino-acid base surfactants, betaine base surfactants, and the like; and non-ionic surfactants such as polyethylene glycol base surfactants, polyalcohol base surfactants, and the like.

[0047] Further, as materials of the cover 6 and the spacer 7 into which the above-mentioned surfactants can be mixed, there are, besides those mentioned above, polybutylene terephthalate, polyamide, polyvinyl chloride, polyvinylidene chloride, polyimide, nylon, and the like.

[0048] As described above, according to the first embodiment of the present invention, the side walls facing the cavity 12 into which blood is drawn, i.e., the portions of the cover 6 and the spacer 7 facing the cavity 12, are made to have hydrophilicity by mixing a chemical having surface activity such as a surfactant or the like into the material itself of the cover 6 and the spacer 7. Therefore, the wettability of the side walls of the cavity 12 is increased, whereby the blood sampled from the suction inlet 8 can be quickly and reliably drawn into the cavity 12. Accordingly, the surfactant layer 11 on the reagent layer 10 can be dispensed with, and the process of manufacturing the biosensor can be simplified.

[0049] The blood suction promoting effect obtained by mixing the surfactant into the insulating base material to be

the cover 6 and the spacer 7 is sufficiently recognized when the surfactant of 0.01 weight % or more is added.

Embodiment 2.

5 [0050] Hereinafter, a second embodiment of the present invention will be described with reference to figure 1.
 [0051] Initially, the construction of a biosensor according to the second embodiment will be described with reference to figure 1. In the first embodiment, a surfactant is mixed into the material itself of the cover 6 and the spacer 7 to make the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity. On the other hand, in this second embodiment, any of the surfactants described for the first embodiment is applied onto an insulating film comprising
 10 polyethylene terephthalate, polycarbonate, or the like and to be a base material of the cover 6 and the spacer 7, or a resin having a hydrophilic polar group at its surface is laminated on the insulating film, so as to coat the insulating film with the surfactant or the resin, thereby making the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity.
 [0052] As the resin having a hydrophilic polar group, there are acrylic resin, polyester resin, urethan resin, and the like.
 15 [0053] Further, when forming the hydrophilic coating on the surface of the insulating base material to be the cover 6 and the spacer 7, the base material is not restricted to the above-mentioned insulating film comprising polyethylene terephthalate or polycarbonate, but other materials such as polybutylene terephthalate, polyamide, polyvinyl chloride, polyvinylidene chloride, polyimide, and nylon may be employed.
 [0054] Furthermore, hydrophilicity of the side walls of the cavity 12 can be increased to enhance wettability of the
 20 side walls by subjecting the surface of the insulating film comprising polyethylene terephthalate, polycarbonate, or the like and to be the base material of the cover 6 and the spacer 7, to primer treatment using organotitanium compound, polyethylene imine compound, isocyanate compound, or the like.
 [0055] As described above, according to the second embodiment, a surfactant is applied onto the insulating film to be the base material of the cover 6 and the spacer 7, or a resin having a hydrophilic polar group at its surface is
 25 laminated on the insulating film so as to coat the surfaces of the cover 6 and the spacer 7 with the surfactant or the resin, whereby the side walls facing the cavity 12 into which blood is drawn, i.e., the portions of the cover 6 and the spacer 7 facing the cavity 12, have hydrophilicity. Therefore, wettability of the side walls of the cavity 12 is increased, whereby the blood sampled from the suction inlet 8 can be quickly and reliably drawn into the cavity 12. Accordingly,
 30 the surfactant layer 11 on the reagent layer 10 is dispensed with, whereby the process of manufacturing the biosensor can be simplified.
 [0056] The blood suction promoting effect is recognized when the thickness of the surfactant layer applied onto the insulating film as the base material of the cover 6 and the spacer 7 or the thickness of the resin layer having a hydrophilic polar radial to be laminated is several tens of angstroms or more. However, in order to sustain the above-mentioned effect for long hours, the thickness is desired to be several hundreds of angstroms or more.

35 Embodiment 3.

[0057] Hereinafter, a third embodiment of the present invention will be described with reference to figure 1.
 [0058] Initially, the construction of a biosensor according to the third embodiment will be described with reference to figure 1. In the first embodiment, a surfactant is mixed into the material itself of the cover 6 and the spacer 7 to make the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity. On the other hand, in this third embodiment, the surfaces of the cover 6 and the spacer 7 facing the cavity 12 are chemically treated or processed to make the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity.
 [0059] As specific methods for chemical surface treatment or processing on the portions of the cover 6 and the spacer 7 facing the cavity 12, there are, for example, corona discharge and glow discharge which are typical plasma discharge processes. In such plasma discharge process, a hydrophilic functional group such as carboxyl group, hydroxyl group, carbonyl group or the like is formed on the surfaces of the cover 6 and the spacer 7 facing the cavity 12, whereby the surface of the material of the cover 6 and the spacer 7 is chemically reformed to increase surface wettability.
 [0060] Further, as materials of the cover 6 and the spacer 7 which can be subjected to the above-mentioned chemical treatment, there are polybutylene terephthalate, polyamide, polyvinyl chloride, polyvinylidene chloride, polyimide, nylon, and the like, in addition to the above-mentioned polyethylene terephthalate and polycarbonate.
 [0061] As described above, according to the third embodiment, the surfaces of the cover 6 and the spacer 7 facing the cavity 12 into which blood is drawn are subjected to the chemical treatment or processing for chemically reforming the surfaces, whereby the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity. Therefore, wettability of the side walls of the cavity 12 is increased, whereby the blood sampled from the suction inlet 8 can be quickly and reliably drawn into the cavity 12. Accordingly, the surfactant layer 11 on the reagent layer 10 is dispensed with, whereby the process of manufacturing the biosensor can be simplified.
 [0062] Further, as processes for chemically reforming the surface property, there are, besides plasma discharge,

coupling reaction, ozone treatment, ultraviolet treatment, and the like, and any of these processes may be employed with the same effects as mentioned above.

Embodiment 4.

5 [0063] Hereinafter, a fourth embodiment of the present invention will be described with reference to figure 1.

[0064] Initially, the construction of a biosensor according to the fourth embodiment will be described with reference to figure 1. In the first embodiment, a surfactant is mixed into the material itself of the cover 6 and the spacer 7 to make the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity. On the other hand, in this fourth 10 embodiment, the surfaces of the cover 6 and the spacer 7 facing the cavity 12 are roughened to form fine and continuous rough-texture (asperities) on the material surface, thereby making the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity.

[0065] As specific methods for roughening the surfaces of the cover 6 and the spacer 7, there are sand blasting, electric discharge, non-glare treatment, mat treatment, chemical plating, and the like. The surfaces of the cover 6 and the spacer 7 facing the cavity 12 are roughened by any of these treatments to increase the surface wettability of the cover 6 and the spacer 7.

[0066] Further, as materials of the cover 6 and the spacer 7 on which such treatment can be performed, there are polybutylene terephthalate, polyamide, polyvinyl chloride, polyvinylidene chloride, polyimide, nylon, and the like, in addition to the above-mentioned polyethylene terephthalate and polycarbonate.

20 [0067] As described above, according to the fourth embodiment, fine and continuous rough-texture (asperities) is formed on the surfaces of the cover 6 and the spacer 7 facing the cavity 12 to make the portions of the cover 6 and the spacer 7 facing the cavity 12 have hydrophilicity. Therefore, wettability of the side walls of the cavity 12 is increased, whereby the blood sampled from the suction inlet 8 can be quickly and reliably drawn into the cavity 12. Accordingly, the surfactant layer 11 on the reagent layer 10 is dispensed with, whereby the process of manufacturing the biosensor 25 is simplified.

Embodiment 5.

30 [0068] Hereinafter, a fifth embodiment of the present invention will be described with reference to figure 1.

[0069] Initially, the construction of a biosensor according to the fifth embodiment will be described with reference to figure 1. In the first to fourth embodiments, the side walls of the cavity 12, i.e., the cover 6 and the spacer 7 facing the cavity 12, are processed so as to have hydrophilicity. In this fifth embodiment, not only the cover 6 and spacer 7 but also the surface of the insulating support 5 on which the working electrode 1, the counter electrode 2, and the reagent layer 10 are formed, are subjected to any of the hydrophilic processes described above.

35 [0070] Hereinafter, a description will be given of the effects obtained by subjecting, not only the cover 6 and the spacer 7, but also the insulating support 5 to the hydrophilic process.

[0071] Initially, as a first effect, when the surface of the insulating support 5 is processed so as to have hydrophilicity, suction of the liquid sample can be further promoted.

40 [0072] For example, in the case where the height of the suction inlet 8 (=the thickness of the spacer 7) is relatively large (0.3mm or more in the sensor shown in figure 1), when the suction inlet 8 sucks, as a liquid sample, blood having a high hematocrit value under a low-temperature environment (10°C or lower), the effect of promoting the suction is not satisfactorily obtained by making only the cover 6 and spacer 7 have hydrophilicity as described above, and the suction ability tends to decrease. So, as well as the cover 6 and the spacer 7, the insulating support 5 is subjected the hydrophilic process as described for any of the first to fourth embodiments, whereby suction of the liquid sample can 45 be further promoted.

[0073] Next, as a second effect, when the electrodes are formed on the surface of the insulating support 5 that has been processed so as to have hydrophilicity, the adhesion of the electrodes to the insulating support 5 is dramatically increased.

50 [0074] For example, in manufacturing biosensors, when a biosensor as shown in figure 1 is obtained by die-cutting an insulating support 5 with a press or the like according to the outline of the sensor after bonding, onto the insulating support 5 on which plural electrodes and reagent layers 10 are formed, a spacer 7 having cut-out grooves for forming cavities 12 in positions corresponding to the respective electrodes and reagent layers, and a cover 6 having air holes 9 in the corresponding positions, the electrodes peel off from the insulating support 5 or the electrodes are cracked due to a shock that occurs when the insulating support 5 is die-cut. This is because the electrodes are formed by 55 printing a paste comprising a conductive material on the insulating support 5 the polarity of which is inherently very small. So, the insulating support 5 is also subjected to the hydrophilic process as described for any of the first to fourth embodiments to make the material surface of the insulating support 5, the surface of which inherently has a very small polarity, have a polarity, whereby spread and adhesion of the paste comprising a conductive material and used as a

material of the electrodes are improved and, therefore, the electrodes are prevented from peeling off from the insulating support 5, or from being cracked.

[0075] As described above, according to the fifth embodiment, since not only the cover 6 and the spacer 7 facing the cavity 12 but also the insulating support 5 are subjected to the hydrophilic process, suction of the blood sampled from the suction inlet 8 is further promoted as compared with the case where only the cover 6 and the spacer 7 are subjected to the hydrophilic process. Furthermore, since the insulating support 5 is subjected to the hydrophilic process before formation of the electrodes to make the insulating support 5 have a polarity, adhesion of the electrodes to the insulating support 5 is increased, whereby peeling-off of the electrodes from the insulating support 5 and cracking of the electrodes, which have occurred during manufacturing of the sensor, are avoided. In the method of roughening the material surface, which is the hydrophilic process described for the fourth embodiment, the level of the rough surface (asperities) at which the effect of adhesion can be expected is within a range of $0.001 \mu\text{m} \sim 1 \mu\text{m}$ and, especially, $0.01 \mu\text{m} \sim 0.1 \mu\text{m}$ are desirable.

[0076] Hereinafter, first and second examples of the present invention will be described.

15 (Example 1)

[0077] On an insulating support 5 which comprises polyethylene terephthalate and has been subjected to corona discharge (power: 400W, rate of discharge: 30m/min), an electrode layer comprising a working electrode 1 and a counter electrode 2 is formed by screen printing, and a reagent layer 10 including an enzyme (glucose oxidase) and an electron acceptor (potassium ferricyanide) is formed on the electrode layer and, thereafter, a spacer 7 comprising polyethylene terephthalate is bonded to a cover 6 comprising polyethylene terephthalate in which about 1% of alkylbenzene sulfonate as an anionic surfactant is blended, thereby fabricating a blood sugar measuring sensor having a groove as a capillary tube into which blood is drawn.

[0078] Table 1 shows the blood suction ability of the sensor so fabricated. Here, a suction inlet 8 having a height of 0.15mm and a width of 2.0mm is used. Each numeric value in Table 1 indicates a time required until the groove as a capillary tube into which blood is drawn is completely filled with the blood, under hostile environments (environmental temperature: 5°C, hematocrit: 65%), and the result proves that the same blood suction promoting effect as that obtained by the conventional sensor is achieved.

30

Table 1

	conventional sensor	sensor of Example 1
1	0.54	0.68
2	0.69	0.58
3	0.69	0.72
4	0.63	0.65
5	0.72	0.64
average(sec)	0.65	0.65
comparison of blood suction rates (n=5)		

35

[0079] While the indices of wettability (surface tension) of the insulating support 5 and the cover 6 which comprise polyethylene terephthalate used in Example 1 are 48dyn/cm when they are not processed, the index of wettability at the surface of the insulating support 5 after being subjected to corona discharge and that at the surface of the cover 6 into which alkylbenzene sulfonate is blended are 54dyn/cm or more, and this result indicates that sufficient wettability for promoting blood suction is secured.

[0080] Figure 2 shows the result of a comparison of the sensor sensitivities at the blood glucose concentrations of 53~992mg/dl. The sensor sensitivity is detected as follows. After the blood is drawn into the capillary tube, a reaction between the reagent and glucose in the blood is promoted for about 25 seconds, and then a voltage of 0.5V is applied between the leads 3 and 4. A current value detected five seconds after the voltage application is the sensor sensitivity. Each numerical value in the graph shown in figure 2 is an average of n=10 times of measuring. As shown in figure 2, the sensitivity of the sensor of Example 1 is about 5% higher than the sensitivity of the conventional sensor. This attests to the result that the disuse of the surfactant layer 11 increases the solubility of the reagent layer 10 that reacts with the blood.

[0081] Table 2 shows the result of a comparison of the repetition accuracy (CV values) in the 10-times measuring. It can be seen from the result in Table 2 that the measuring variations in the sensor of Example 1 (variations in each

sensor) are significantly reduced as compared with the measuring variations in the conventional sensor.

Table 2

glucose concentration	conventional sensor	sensor of Example 1
53mg/dl	6.25%	3.79%
83mg/dl	3.15%	1.67%
253mg/dl	3.49%	1.53%
488mg/dl	2.24%	0.60%
596mg/dl	2.49%	1.86%
992mg/dl	2.23%	2.11%
comparison of sensor accuracy (CV values)		

[0082] As is evident from the results of figure 2 and table 2, a highly-sensitive biosensor with less variations can be realized by employing the sensor of Example 1.

[0083] Further, it is also confirmed how much the adhesion between the electrode layer and the insulating support 5 is improved by subjecting the surface of the insulating support 5 to corona discharge. A checker pattern having 100 squares at 1mm intervals is formed according to JISK5400 (general test method for coating; adhesion; checker-pattern taping method), and the degree of electrode peeling-off is checked with an adhesive cellophane tape. The result is as follows. While peeling-off of electrodes occurs at frequency of 5/100 squares in the conventional sensor performing no corona discharge, it occurs at frequency of 0/100 squares in the sensor of Example 1, that is, a clearly significant difference is confirmed.

(Example 2)

[0084] On an insulating support 5 comprising polyethylene terephthalate, an electrode layer comprising a working electrode 1 and a counter electrode 2 is formed by screen printing, and a reagent layer 10 including an enzyme (glucose oxidase) and an electron acceptor (potassium ferricyanide) is formed on the electrode layer and, thereafter, a spacer 7 comprising polyethylene terephthalate is bonded to a cover 6 comprising a compound film (the index of surface wettability: 54dyn/cm or more) which is obtained by laminating a polyester base resin having a hydrophilic polar group on polyethylene terephthalate, thereby fabricating a blood sugar measuring sensor having a groove as a capillary tube into which blood is drawn, and evaluations similar to those of Example 1 are executed. Table 3 shows the result of a comparison of the blood suction rates between the sensor fabricated as described above and the conventional sensor, figure 3 shows the result of a comparison of the sensor sensitivities at the blood glucose concentrations of 53~992mg/dl, and Table 4 shows the result of a comparison of the repetition sensor accuracy (CV values) in 10-times measuring.

Table 3

	conventional sensor	sensor of Example 2
1	0.54	0.62
2	0.69	0.55
3	0.69	0.68
4	0.63	0.60
5	0.72	0.69
average(sec)	0.65	0.63
comparison of blood suction rates (n=5)		

Table 4

glucose concentration	conventional sensor	sensor of Example 2
53mg/dl	6.25%	3.88%
83mg/dl	3.15%	2.17%

Table 4 (continued)

glucose concentration	conventional sensor	sensor of Example 2
253mg/dl	3.49%	1.22%
488mg/dl	2.24%	1.60%
596mg/dl	2.49%	1.56%
992mg/dl	2.23%	2.05%
comparison of sensor accuracy (CV values)		

[0085] From these results, excellent blood suction ability and sensor responsivity (sensitivity, CV value) as high as those of Example 1 are confirmed.

15 APPLICABILITY IN INDUSTRY

[0086] A biosensor according to the present invention is available as a biosensor which improves sensitivity and reduces variations when analyzing a specific component in a liquid sample which is drawn into a cavity of the sensor by capillary phenomenon.

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Claims

1. A biosensor which is provided with a cavity into which a liquid sample is drawn by capillary phenomenon, and is able to analyze a component in the liquid sample by a reaction between the drawn liquid sample and a reagent, wherein
 - the surface itself of at least a portion of side walls of the sensor, said side walls facing the cavity, has hydrophilicity.
2. A biosensor as defined in Claim 1 wherein
 - the side walls of the sensor facing the cavity are made of a resin material in which a surfactant is mixed.
3. A biosensor as defined in Claim 2 wherein
 - the amount of the surfactant to be mixed is 0.01 weight % or more.
- 35 4. A biosensor as defined in Claim 1 wherein
 - the side walls of the sensor facing the cavity are made of a film the surface of which is covered with a surfactant.
5. A biosensor as defined in Claim 1 wherein
 - the side walls of the sensor facing the cavity are made of a film the surface of which is covered with a resin having a hydrophilic polar group.
- 40 6. A biosensor as defined in Claim 4 or 5 wherein
 - the thickness of the surfactant or the resin having a hydrophilic polar group, which covers the film, is several tens of angstroms or more.
7. A biosensor as defined in Claim 1 wherein
 - the surface of at least a portion of the side walls forming the cavity is chemically reformed.
- 50 8. A biosensor as defined in Claim 7 wherein
 - a hydrophilic functional group is formed on the surface of at least a portion of the side walls facing the cavity, by subjecting the surface to any of the following treatments: plasma discharge, coupling reaction, ozone treatment, and UV treatment.
- 55 9. A biosensor as defined in Claim 1 wherein
 - the surface of at least a portion of the side walls facing the cavity is made of a rough surface.

10. A biosensor as defined in Claim 9 wherein
a rough surface is formed at the surface of at least a portion of the side walls facing the cavity, by subjecting
the surface to any of the following treatments: sand blasting, electric discharge, non-glare treatment, mat treatment,
and chemical plating.

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11. A biosensor as defined in any of Claims 1 to 10 wherein
the surface of the side wall, on which the reagent that reacts with the liquid sample is formed, has hydrophilicity.

10 12. A biosensor as defined in any of Claims 1 to 10 wherein
the surface of the side wall, on which electrodes that detect the reaction between the liquid sample and the
reagent are formed, has hydrophilicity.

15 13. A biosensor as defined in Claim 12 wherein
the surface of the support is made of a rough surface, and the level of the rough surface to be formed is
0.001 μ m to 1 μ m.

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Fig.1

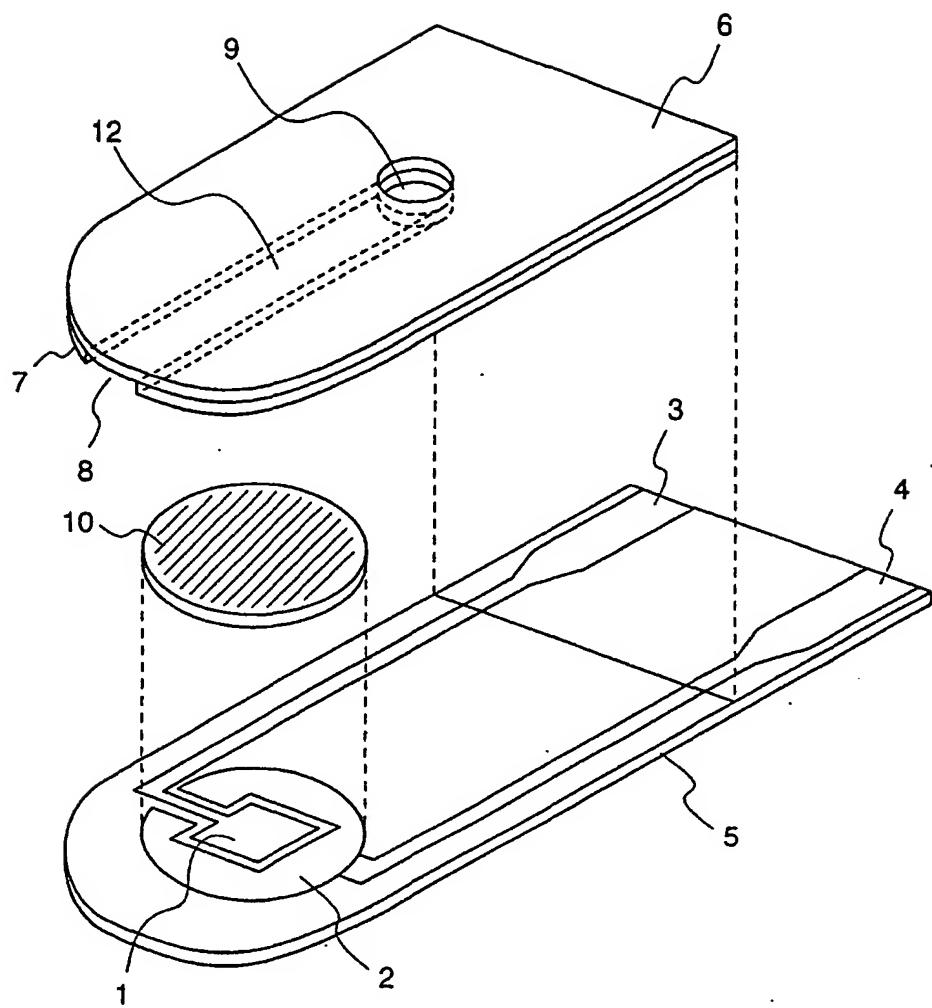


Fig.2

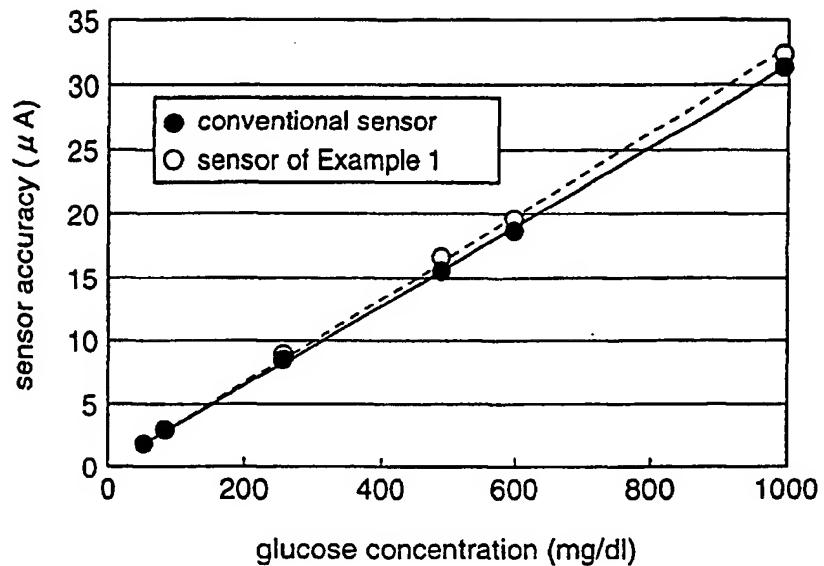


Fig.3

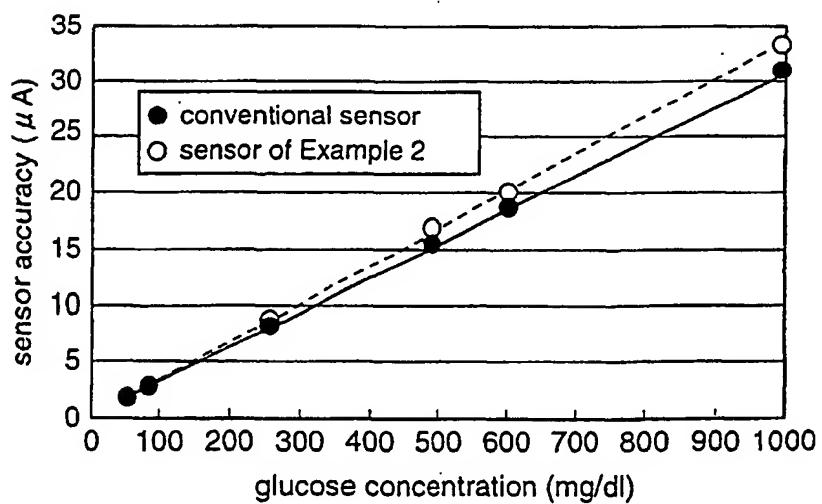
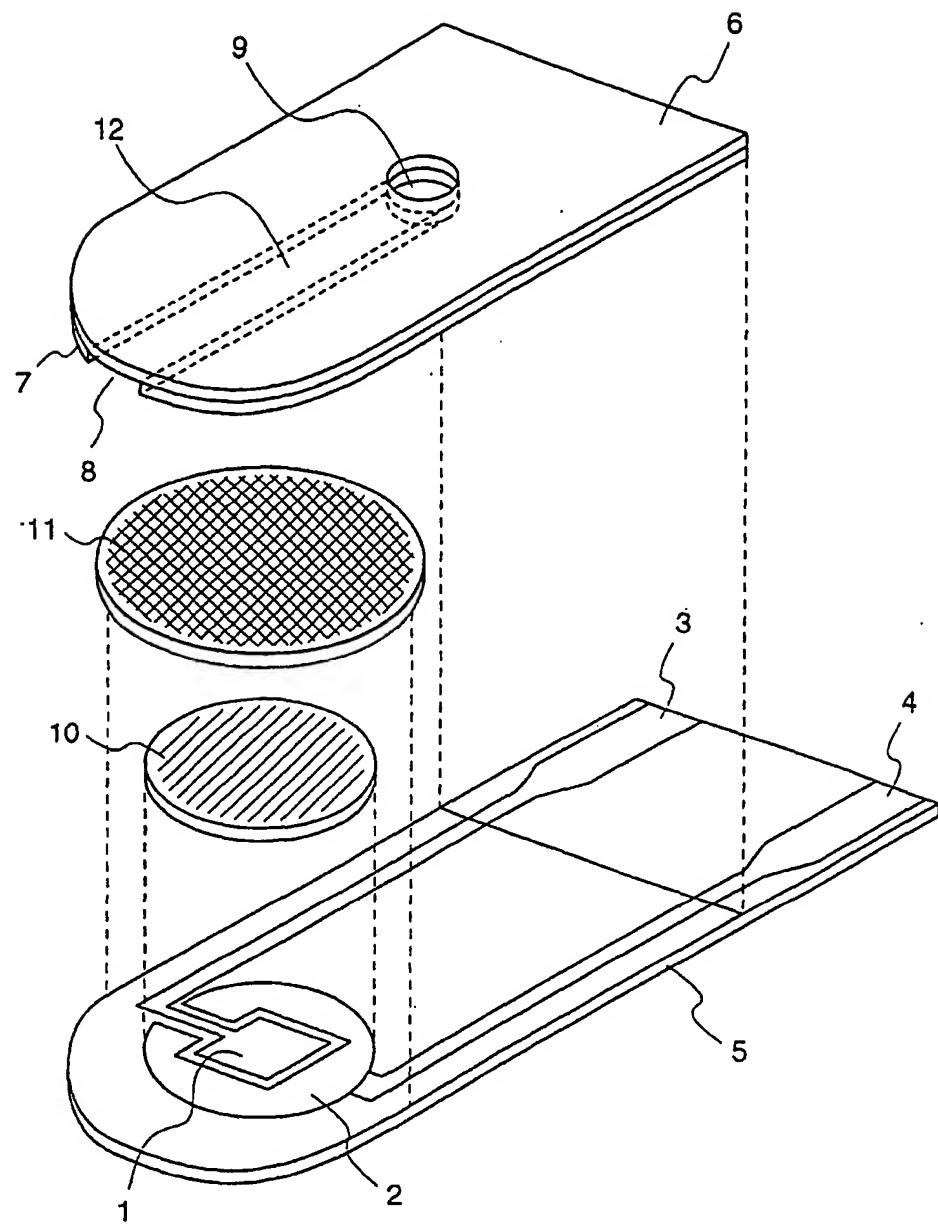


Fig.4



INTERNATIONAL SEARCH REPORT		International application No. PCT/US00/33629
A. CLASSIFICATION OF SUBJECT MATTER IPC(7) :Please See Extra Sheet. US CL : 530/327, 328, 326, 325, 324; 514/12, 13, 14, 15; 536/23.1 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 530/327, 328, 326, 325, 324; 514/12, 13, 14, 15; 536/23.1		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) WEST 2.0, MEDLINE, BIOSIS, EMBASE search terms: author names, p53, peptide?, hla		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 96/03140 A1 (CYTEL CORPORATION) 08 February 1996, see entire document.	1-34
Y	WO 97/34617 A1 (CYTEL CORPORATION) 25 September 1997, see entire document.	1-34
Y	WO 94/03205 A1 (CYTEL CORPORATION) 17 February 1994, see entire document.	1-34
<input type="checkbox"/> Further documents are listed in the continuation of Box C.		<input type="checkbox"/> See patent family annex.
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T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art *&* document member of the same patent family		
Date of the actual completion of the international search 05 FEBRUARY 2001		Date of mailing of the international search report 16 MAR 2001
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230		Authorized officer  RON SCHWADRON Telephone No. (703) 308-0196

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A. CLASSIFICATION OF SUBJECT MATTER:
IPC (7):

A61K 38/00, 38/03, 38/08, 38/16, 39/00; C07H 21/00; C07K 9/00, 14/00